

# Generation of gene-engineered T cells

# **T cell transduction process**

## **Application**

Fully automated labelling, enrichment, activation, transduction and expansion of human T cells from patient material for production of gene-engineered T cells.

This application sheet gives an overview of the specifications and material to perform the T cell transduction (TCT) process. Furthermore, it provides an overview of the setup for the tubing set, general workflow and performance data.

## **Specifications**

Process name:	T cell transduction process			
Selection capacity:	up to 3×10 <sup>9</sup> cells			
Sample volume for selection:	50–280 mL			
TransAct™ stimulation capacity:	1×10 <sup>8</sup> cells (0.2–2×10 <sup>8</sup> )			
Expansion capacity:	maximum 2×10 <sup>7</sup> cells/mL			
Final product elution volume:	100 mL			
Process time:	8–14 days			

#### **Products**

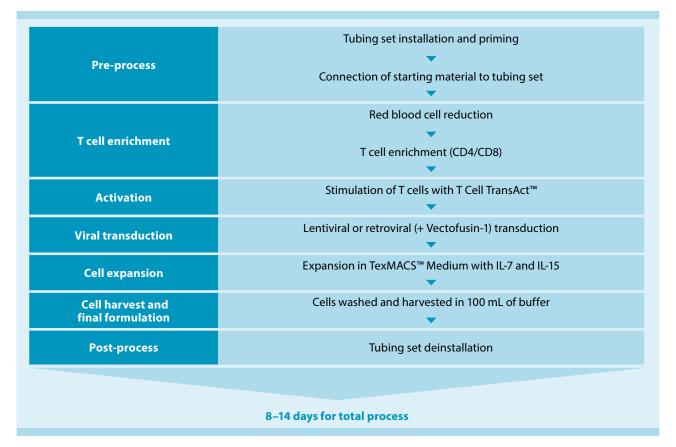
Consumables	Amount required
CliniMACS Prodigy®	1 piece
CliniMACS Prodigy TS 520	1 set
CliniMACS® PBS/EDTA Buffer	1 bag
TexMACS <sup>™</sup> GMP Medium	3×2 L bags
CliniMACS CD4 Reagent	1 vial
CliniMACS CD8 Reagent	1 vial
MACS® GMP T Cell TransAct	1 vial
MACS GMP Recombinant Human IL-7	3 vials
MACS GMP Recombinant Human IL-15	3 vials

Additional materials	Amount required
Triple sampling adaptor	1 piece
Transfer bag 150 mL	1 bag
Luer/Spike Interconnector	1 piece
MACS GMP Vectofusin-1*	1 vial
Transfer bag 1000 mL	1 bag

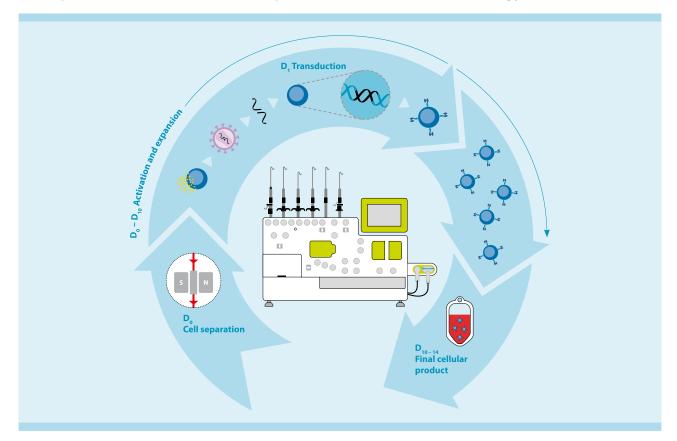
Additional equipment and materials		
Sterile docking device		
Cell counter		
Flow cytometer		
Syringes and hypodermic needles		
Human serum albumin		
Final formulation buffer		
Human AB serum		

\*Please discuss your specific requirements with your Miltenyi Biotec representative.

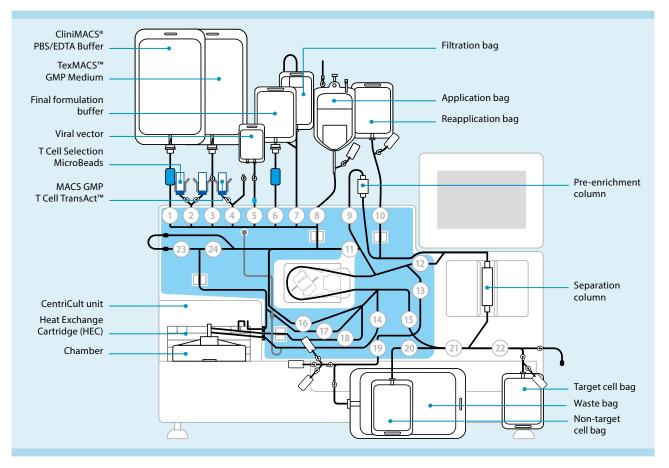
#### **Process overview**



## Principle of the T cell transduction process on the CliniMACS Prodigy®



# CliniMACS Prodigy® TS 520 setup



## **Performance data**

		Final product					
Starting product CD4 <sup>+</sup> and CD8 <sup>+</sup> T cells (%)	Isolated CD4 <sup>+</sup> and CD8 <sup>+</sup> T cells (%)	CD4 <sup>+</sup> and CD8 <sup>+</sup> T cells (%)	CAR <sup>+</sup> T cells (%)	Viability (%)	CAR⁺ T cell number (×10°)		
Performance data of healthy donor or patient derived material							
72%±11%	83%±9%	91%±5%	34%±12%	90%±4%	1.4±0.7		
23%±21%	55%±9%	88%±7%	36%±18%	88%±7%	1.0±0.4		
Performance data of healthy donor with or without human AB serum (3%) for culture							
58%±17%	82%±14%	92%±3%	36%±9%	93%±3%	1.6±0.3		
60%±15%	87%±7%	94%±6%	43%±12%	93%±3%	2.9±1.3		
	and CD8 <sup>+</sup> T cells (%) lata of healthy donor or p 72%±11% 23%±21% lata of healthy donor with 58%±17%	and CD8* T cells (%)CD8* T cells (%)lata of healthy donor or patient derived mate72%±11%83%±9%23%±21%55%±9%lata of healthy donor with or without human58%±17%82%±14%	and CD8* T cells (%)     CD8* T cells (%)     CD8* T cells (%)       ata of healthy donor or patient derived material       72%±11%     83%±9%     91%±5%       23%±21%     55%±9%     88%±7%       ata of healthy donor with or without human AB serum (3%) for     58%±17%     92%±3%	Starting product CD4*     Isolated CD4* and CD8* T cells (%)     CD4* and CD8* T cells (%)     CAR* T cells (%)       Iata of healthy donor or patient derived material     83%±9%     91%±5%     34%±12%       23%±21%     55%±9%     88%±7%     36%±18%       Iata of healthy donor without human AB serum (3%) for culture     58%±17%     36%±9%	Starting product CD4+ and CD8+T cells (%)     Isolated CD4+ and CD8+T cells (%)     CDA+ and CD8+T cells (%)     CAR+T cells (%)     Viability (%)       Iata of healthy donor or patient derived material     83%±9%     91%±5%     34%±12%     90%±4%       23%±21%     55%±9%     88%±7%     36%±18%     88%±7%       Iata of healthy donor without human AB serum (3%) for ulture     93%±3%		

#### References

Lock, D. and Mockel-Tenbrinck, T. *et al.* (2017) Human Gene Therapy 28(10): 914–925.
Mock, U. *et al.* (2016) Cytotherapy 18(8): 1002–11.



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